



Metabolism
Clinical and Experimental

Metabolism Clinical and Experimental 57 (2008) 1563-1569

www.metabolismjournal.com

Plasma basic fibroblast growth factor is correlated with plasminogen activator inhibitor—1 concentration in adults from the Veterans Affairs Diabetes Trial

Mark B. Zimering^{a,b,*}, Robert J. Anderson^{c,d}, Ping Luo^c, Thomas E. Moritz^c Investigators for the Veterans Affairs Diabetes Trial^c

^aMedical Service, Department of Veterans Affairs New Jersey Health Care System, Lyons, NJ 07939, USA
 ^bUniversity of Medicine and Dentistry of New Jersey, Robert Wood Johnson Medical School, New Brunswick, NJ, USA
 ^cHines Cooperative Studies Program Coordinating Center, Veterans Affairs Hospital, Hines, IL, USA
 ^dDivision of Epidemiology and Biostatistics, School of Public Health, University of Illinois at Chicago, Chicago, IL, USA
 Received 7 February 2008; accepted 16 June 2008

Abstract

Basic fibroblast growth factor (bFGF) is a potent mitogen in endothelial and vascular smooth muscle cells that increases in serum from adults with coronary artery disease and in microalbuminuric type 2 diabetes mellitus. There has been no prior study of plasma bFGF as a possible cardiovascular risk marker in type 2 diabetes mellitus. In this study, we tested for a correlation between log plasma bFGF immunoreactivity (bFGF-IR) and baseline cardiovascular risk factors in a baseline subset of subjects with advanced type 2 diabetes mellitus from the Veterans Affairs Diabetes Trial ([mean] age, 60 years; hemoglobin A_{1c} , 9.5%; diabetes' duration, 11 years). Plasma bFGF-IR was determined with a sensitive, specific, 2-site enzyme-linked immunoassay in 281 patients at the baseline visit. Results were compared with baseline risk factors or baseline medication use. Baseline plasma bFGF-IR ranged from 0 to 141 pg/mL. Log plasma bFGF correlated significantly with non-Hispanic white race (P = .002), waist-hip ratio (P = .002), and plasminogen activator inhibitor–1 concentration (P < .0001). Log plasma bFGF correlated inversely with African American race (P = .0003). In multiple regression analysis, plasminogen activator inhibitor–1 and race were significantly correlated with log plasma bFGF. These results suggest a significant correlation between log plasma bFGF-IR and plasminogen activator inhibitor–1, a marker of hemostatic risk. Published by Elsevier Inc.

1. Introduction

Cardiovascular disease is the leading cause of mortality in adult type 2 diabetes mellitus. Basic fibroblast growth factor (bFGF) is a potent angiogenesis factor and broad-spectrum cell mitogen in endothelial, epithelial, and mesenchymalderived cells [1]. Plasma bFGF is low or undetectable in healthy subjects [2], but increases in those with coronary artery disease [3,4]. Basic FGF may function as a local vascular growth factor to promote atherosclerosis [5,6].

E-mail address: mark.zimering@med.va.gov (M.B. Zimering).

Veterans Affairs Diabetes Trial (VADT) is a large ongoing clinical trial in adult patients with type 2 diabetes mellitus randomized to standard or intensive glycemic control [7]. The ongoing VADT substudy will test whether plasma bFGF is a novel marker of prospective (5 years) cardiovascular risk in adults with type 2 diabetes mellitus. The present baseline report analyzed the relationship between baseline plasma bFGF and baseline risk factors.

We now report that plasma bFGF is substantially increased in a subset of adult men with long-standing type 2 diabetes mellitus. Plasma bFGF levels varied widely from 0 to 141 pg/mL in 281 subjects. Because of the well-known log-linear dose-response relationship in bFGF-induced cellular proliferation over a similar range of bFGF concentrations (1-100 pg/mL) [1,8], a natural log transformation of bFGF values was performed to optimize the ability

^{*} Corresponding author. Department of Veterans Affairs New Jersey Health Care System, Medical Service, Lyons, NJ 07939, USA. Tel.: +1 908 647 0180x4426; fax: +1 908 604 5249.

to detect biologically significant correlations between bFGF and established cardiovascular risk factors.

Log plasma bFGF directly correlated with non-Hispanic white race, waist-hip ratio, and plasma plasminogen activator inhibitor—1 (PAI-1) level and inversely with African American race. These are the first in vivo data suggesting a correlation between log plasma bFGF and PAI-1; the latter is an important hemostatic risk marker.

2. Subjects and methods

2.1. Study subjects

Informed consent for the Investigational Review Board–approved substudy was obtained from 281 diabetic subjects at 5 outpatient sites who had consented to participate in the main VADT. Plasma was prepared from the EDTA-anticoagulated blood drawn in the morning from fasting subjects at each site. Plasma was aliquoted and shipped frozen (dry ice) to a central laboratory (Maveric, Boston Veterans Affairs Medical Center, Boston, MA), where it was inventoried and stored at –80°C for 1 to 2 years. Archived, coded, and frozen EDTA plasma from consecutively enrolled patients was shipped to Dr Zimering's laboratory (VA New Jersey Health Care System, Lyons, NJ), where bFGF immunoreactivity (bFGF-IR) assays were performed. All other assays were performed at the Central Laboratory of the VADT (Tufts University, Boston, MA).

2.2. Baseline characteristics

Baseline clinical characteristics are summarized in Table 1. Plasma bFGF-IR was determined at the baseline study visit. All subjects were older than 40 years. Ninety-

Table 1 Baseline characteristics in study subjects

	Mean	SD
n	281	
bFGF (pg/mL)	11.5	16.7
log bFGF ^a	1.34	1.93
Age (y)	59.6	8.5
BMI (kg/m^2)	31.1	4.8
Waist circumference (cm)	108.9	12.9
Hip circumference (cm)	109.3	9.5
SBP (mm Hg)	130.9	18.1
DBP (mm Hg)	74.3	10.6
Diabetes' duration (y)	11.5	7.6
Urine albumin-creatinine ratio (mg/g)	138	422
HbA _{1c} (%)	9.5	1.4
Triglyceride (mg/dL)	193	190
Total cholesterol (mg/dL)	182	42
LDL cholesterol (mg/dL)	107	33
HDL cholesterol (mg/dL)	37	9

BMI indicates body mass index; SBP, systolic blood pressure; DBP, diastolic blood pressure.

seven percent of subjects were male; 16%, Hispanic; 20%, African American; and 63%, non-Hispanic white. Fourteen percent reported baseline occurrence of myocardial infarction (MI). Patients with chronic kidney disease were excluded from study participation.

2.3. Medications

All patients were taking antidiabetic medications at baseline including oral agents and/or insulin. Baseline antihypertensive medication use included a thiazide diuretic (15%), calcium channel blocker (20%), angiotensin-converting enzyme (ACE) inhibitor (64%), and angiotensin II receptor blocker (ARB) (5.5%). Sixty-two percent of patients used a statin at baseline, 16% reported fibrate drug use, and 76% reported daily aspirin use. Twenty-one percent of patients used a cyclooxygenase inhibitor drug; nearly all such drugs belonged to the nonsteroidal anti-inflammatory class of medications.

2.4. Laboratory and clinical measures

Urinary microalbumin, plasma glycosylated hemoglobin (HbA_{1c}), and urine creatinine were determined by standard methods as previously described [7]. Urinary albumincreatinine ratio was calculated as albumin concentration/ creatinine concentration ×100. Normo-, micro-, and macroalbuminuria are defined as albumin-creatinine ratio of less than 30, 30.1 to 300, and at least 300.5 mg/g, respectively. Plasma total cholesterol, triglycerides, and high-density lipoprotein (HDL) cholesterol were determined by standardized direct enzymatic assay methods as previously reported [7]. Low-density lipoprotein (LDL) cholesterol was calculated using the Friedewald equation on all samples with plasma triglyceride concentration less than 400 mg/dL. Blood pressure was recorded in the seated position after 5-minute rest. Three consecutive readings were obtained, and the median value of the 3 consecutive determinations was used for analysis.

2.5. Plasma samples

Archived, coded EDTA plasma samples were kept frozen (-40°C) for up to 2 years before assay for bFGF-IR. Plasma bFGF-IR and bFGF-like bioactivity were previously shown to be stable for 5 years or longer at -20°C, and for up to 3 freeze-thaw cycles [9].

2.6. Basic fibroblast growth factor assays

Basic FGF-IR in plasma was determined using a sensitive, specific, 2-site enzyme-linked immunoassay (R&D Systems, Minneapolis, MN).

The mean minimal detectable dose of FGF-2 was 0.5 pg/mL (n = 9 assays). The method was linear between 0.5 and 64 pg/mL. The average correlation coefficient (between bFGF dose and absorbance) for the runs was 0.99. The intraassay coefficients of variation for low- and high-dose calibration standards or human diabetic plasma samples

^a Results are natural log (bFGF + 0.05), such that bFGF values less than the assay detection limit, that is, 0, do not result in log bFGF that is undefined

were less than or equal to 8%; the interassay coefficients of variation for patient samples or calibration standards ranged from 10% to 14%. Recovery of bFGF-IR in diluted (1:2) samples of normal human plasma ranged from 108% to 123%. The dilution curves of patient plasma samples were parallel to the standard curve. Fibroblast growth factor–1, FGF-4 (hst), FGF-5, and FGF-6 did not cross-react in the assay. In prior studies that used the same bFGF-IR assay method, mean serum bFGF-IR in 15 healthy subjects (men and women; age range, 39-74 years) was 0.9 pg/mL (range, 0-4 pg/mL) [10].

Plasma bFGF-IR in 43 healthy male blood donors, aged 21 to 63 years, ranged from 0 to 4 pg/mL; and there was no effect of age on plasma bFGF level [11].

2.7. Statistics

Basic FGF-IR was not normally distributed, at least partially because of a number of below-detection-limit

Table 2
Relation of log bFGF to baseline categorical risk factors

Categorical variables	Mean of log (bFGF + 0.05)		P value ^a	
	Yes	No		
Demographics				
Sex (M/F)	1.34	1.33	.9601	
Hispanic (Y/N)	1.58	0.92	.8111	
Non-Hispanic white (Y/N)	1.49	1.31	.0019	
Black (Y/N)	0.47	1.55	.0003	
Current smoker (Y/N)	1.22	1.36	.6053	
Exercise regularly (Y/N)	1.52	1.21	.3366	
Drug use at baseline				
Aspirin (Y/N)	1.38	1.19	.8051	
ACE inhibitor (Y/N)	1.31	1.38	.9267	
ARB (Y/N)	0.12	1.41	.0726	
Thiazide diuretic (Y/N)	1.80	1.25	.0545	
Statin (Y/N)	1.28	1.43	.8428	
Fibrate (Y/N)	1.38	1.33	.8518	
Calcium channel antagonist (Y/N)	0.85	1.46	.0561	
Thiazolidinedione (Y/N)	1.53	1.28	.3653	
Insulin (Y/N)	1.26	1.40	.7402	
Sulfonylurea (Y/N)	1.41	1.20	.6624	
Metformin (Y/N)	1.30	1.42	.4418	
Gabapentin (Y/N)	1.98	1.30	.1764	
Aldactone (Y/N)	2.20	1.33	.4430	
α-Adrenoceptor blocker (Y/N)	1.37	1.33	.6717	
COX inhibitor (Y/N)	1.19	1.38	.8267	
Furosemide (Y/N)	1.49	1.32	.9314	
History				
Hypertension (Y/N)	1.38	1.24	.1878	
MI (Y/N)	1.74	1.27	.2339	
Coronary revascularization (Y/N)	1.21	1.37	.1643	
Stroke (Y/N)	1.24	1.34	.9795	
Albuminuria ^b				
Macroalbuminuria	1.03		.4616	
Microalbuminuria	1.29			
Normoalbuminuria	1.42			

Log indicates natural log; Y, presence (bolded) of indicated risk factor; COX, cyclooxygenase.

Table 3
Relation of log bFGF to baseline continuous risk factors

Continuous variables	Spearman correlation		
	Coefficients	P value ^a	
Age	0.0574	.3378	
BMI	0.0009	.9875	
Waist circumference	0.0356	.5599	
Hip circumference	-0.1097	.0714	
Average SBP	-0.0135	.8236	
Average DBP	0.0022	.9705	
Total cholesterol	-0.0038	.9500	
LDL cholesterol	-0.0219	.7242	
HDL cholesterol	-0.0246	.6833	
Triglycerides	-0.0025	.9667	
Urine albumin-creatinine ratio	-0.0358	.5528	
Plasma fibrinogen	0.0606	.3178	
PAI-1	0.2820	<.0001	
Waist-hip ratio	0.1857	.0021	
Duration of diabetes	-0.0481	.4218	
HbA _{1c}	-0.1272	.0331	
Fasting plasma glucose	-0.0752	.2095	
Serum creatinine	-0.1863	.3332	

 $^{^{\}mathrm{a}}$ P value for correlation with log (bFGF + 0.05), where "log" is the natural log.

values and partially because of extremely high values. The Wilcoxon rank sum test was used for group comparisons of bFGF-IR (Table 3), and the correlations reported are Spearman correlation coefficients.

3. Results

3.1. Relation of log bFGF to baseline characteristics

Baseline log plasma bFGF was directly correlated with non-Hispanic white race (P = .002) and inversely with African American race (P = .0003) (Table 2). Baseline log plasma bFGF was directly correlated with plasma PAI-1 (P < .0001) and waist-hip ratio (P = .002), and was inversely

Table 4
Multiple regression analyses of log bFGF and covariates including (A) non-Hispanic white or (B) African American race

Variable	Parameter	SE	P value
	estimate		
\overline{A}			
Intercept	1.005	1.975	.61
PAI-1	0.013	0.005	.01
Waist-hip ratio	1.623	1.843	.38
HbA _{1c}	-0.244	0.094	.01
NHW	0.525	0.259	.04
NHW indicates non-His	spanic white.		
В			
Intercept	1.852	1.973	.35
PAI-1	0.011	0.005	.03
Waist-hip ratio	1.413	1.832	.44
HbA _{1c}	-0.251	0.093	.008
African American n = 236 subjects	-0.845	0.307	.006

^a P value based on χ^2 statistic.

^b P value from Fisher exact test.

Table 5
Prevalence of baseline cardiovascular diseases by ethnicity

	Non-Hispanic white	Hispanic or African American	P value
Baseline MI	32 (19.1%)	7 (7.6%)	.014
Any macrovascular event	77 (45.8%)	25 (27.2%)	.003

Macrovascular event includes MI, stroke, angina, coronary artery bypass, percutaneous coronary intervention, and peripheral vascular disease.

correlated with HbA_{1c} (P=.03) (Table 3). No significant correlation was identified between log plasma bFGF-IR and patient age, diabetes' duration, or other continuous baseline risk factors (Table 3); and no significant difference in mean log plasma bFGF-IR was detected when comparing groups defined by baseline presence of MI, cigarette smoking, stroke, or hypertension (Table 2).

3.2. Relation of bFGF-IR to baseline medication use

Mean baseline log plasma bFGF-IR was higher with baseline thiazide diuretic use (P = .05) (Table 2) and lower at borderline significance with baseline ARB (P = .07) or calcium channel antagonist use (P = .06) (Table 2). There was no detected difference of mean plasma bFGF-IR with use of insulin, metformin, sulfonylurea, or a thiazolidine-dione (Table 2).

3.3. Independent correlates of log plasma bFGF

In multiple regression analyses, PAI-1 (P = .01) and non-Hispanic white race (P = .04) were significantly correlated with log plasma bFGF-IR (Table 4A). African American race (P = .006) and baseline HbA_{1c} (P = .008) were inversely correlated with log plasma bFGF (Table 4B).

3.4. Prevalence of cardiovascular diseases by ethnicity

Baseline MI was reported in a significantly higher proportion of non-Hispanic white subjects (19.1%) com-

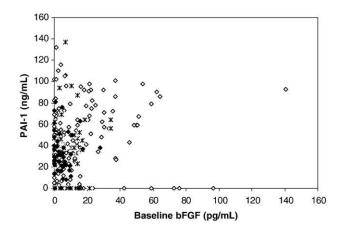
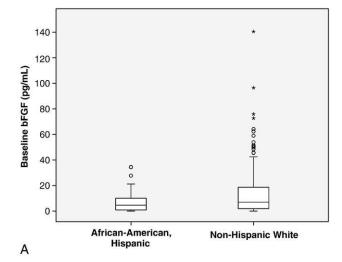


Fig. 1. Scatter plot of PAI-1 vs baseline plasma bFGF values (open diamonds represent non-Hispanic white subjects, stars represent Hispanic subjects, and filled diamonds represent African American subjects).



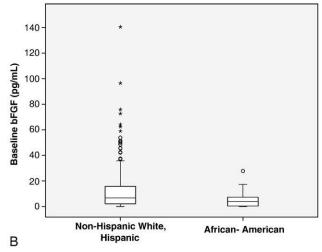


Fig. 2. Box/whisker plot of bFGF values in (A) non-Hispanic white or (B) African American diabetic subjects compared with subjects of other race/ethnicity. Lines inside boxes illustrate the median value; 25th and 75th percentile cutoff values: the lower and upper edges of the box. Whiskers illustrate the range of values occurring within a distance = 1.5 times the interquartile range (IQR) from the lower or upper edge of the box. Outliers are values between 1.5 and 3 times the IQR from the upper edge of the box and are represented by open circles; extreme values more than 3 times the IQR from the upper edge of the box are represented by asterisks.

pared with Hispanic or African American subjects (7.6%, P = .01 for the difference; Table 5). Any prior macrovascular event was reported in a significantly higher proportion of non-Hispanic white subjects (45.8%) compared with events in Hispanic or African American subjects (27.2%, P = .003 for the difference; Table 5).

3.5. Distribution of bFGF and PAI-1 values

Baseline plasma bFGF showed a wide range in values, from 0 to 141 pg/mL (Fig. 1). All but a few bFGF values greater than 20 pg/mL were observed in non-Hispanic white patients (Fig. 2A, B). Exponentially increased bFGF values (≥20 pg/mL) were generally associated with higher values of PAI-1 (Fig. 1). The mean PAI-1 level among

African American subjects (33.2 \pm 17.6 ng/mL) was significantly lower compared with the mean PAI-1 levels in non-Hispanic white (50.5 \pm 28.7 ng/mL) or in Hispanic subjects (50.1 \pm 23.1 ng/mL, P = .0004 for the comparison) (data shown in Fig. 1).

4. Discussion

Circulating bFGF increases in a wide spectrum of cancers [12,13], consistent with its role as a tumor angiogenesis factor [14]. Yet there have been few if any reports of plasma bFGF levels from a large group of adults with long-standing type 2 diabetes mellitus. No prior study demonstrated both striking elevation [9] and wide variability in bFGF levels as were observed in the present study.

Basic FGF is one of the most potent known angiogenic factors. Half-maximal doses of bFGF needed for proliferative activity in many different cell types (10-50 pg/mL) [1] approximate the levels observed in a substantial proportion of our diabetic subjects (Fig. 1). Whether such high bFGF levels are transient or may persist in the circulation could not be determined in this study. The independent significant correlation between log plasma bFGF and PAI-1 may be consistent with bFGF-induced synthesis of PAI-1 reported in vitro [15,16]. It suggests that substantially elevated plasma bFGF is likely to be biologically active rather than an assay artifact. Basic FGF could directly contribute to PAI-1 levels in vivo or be tightly associated with one or more additional factors that induce both PAI-1 and bFGF synthesis.

Plasminogen activator inhibitor-1 inhibits fibrinolysis, resulting in a prothrombotic state. Plasminogen activator inhibitor-1 levels increase in diabetes [17]; and in prior studies, high PAI-1 level was associated with an increased risk for first or recurrent MI [18,19]. Plasminogen activator inhibitor-1 levels vary by race/ethnicity: African American subjects had the lowest levels in a multiethnic study of hemostatic markers [20], and there was evidence for decreased endothelial cell expression of PAI-1 among African American subjects [21]. Our findings of substantially lower log plasma bFGF and lower plasma PAI-1 among African American diabetic subjects may be consistent with common factors regulating the local endothelial production of both PAI-1 and bFGF. Additional factors, some perhaps more closely related to body mass index than was bFGF, for example, transforming growth factor- β [22], may have contributed to a subset of high plasma PAI-1 despite low plasma bFGF (Fig. 1). Markedly elevated plasma bFGF may not only promote angiogenesis, smooth muscle cell proliferation [23], and tumor cell proliferation [1]; but log bFGF may indicate (through its significant association with PAI-1) an increased risk for thrombosis in subsets of obese patients with type 2 diabetes mellitus.

The tissue sources for exponentially increased plasma bFGF are unknown. Human omental tissue is highly vascular: it contained much higher concentrations of a bioactive bFGF-like protein compared with normal or neoplastic renal or prostate tissues [24,25]. Omental preadipocytes from obese humans also expressed much higher levels of bFGF messenger RNA compared with those from lean control subjects [26]. Whether bFGF is released into the general circulation from omental storage sites is unknown. One possible mechanism for its release may involve macrophages abundant in visceral adipose tissue [27] that can release proteases capable of liberating bFGF from storage sites in extracellular matrix [28].

Angiotensin II increases vascular smooth muscle cell bFGF synthesis [29] and may promote increased plasma bFGF levels in subsets of diabetic patients. In a prior study in adults with microalbuminuric type 2 diabetes mellitus, treatment with ACE inhibitor drugs was associated with substantially lower levels of plasma bFGF bioactivity or immunoreactivity [9]. Because plasma renin activity varies considerably in humans [30], the present findings of substantially lower mean log plasma bFGF in African American subjects may be consistent (in part) with low renin hypertension common among African American patients [31]. Lower mean log bFGF for ARB use (Table 2) is consistent with a role for angiotensin II in the elaboration of increased plasma bFGF. Associations between log bFGF and other medications must be interpreted cautiously, however, because there may have been confounding by race (eg, increased use of calcium channel antagonists among African American hypertensive patients).

Other unknown factors may have contributed to substantially lower plasma bFGF among African American diabetic patients. It is also unclear whether inflammation, insulin resistance, or chance may have contributed to the apparent direct correlation between non-Hispanic white race and log plasma bFGF. For example, proinflammatory cytokines (tumor necrosis factor— α , interleukin-1, and interferon- γ) associated with visceral obesity [32] induced the synthesis and release of high levels of bFGF from normal endothelial cells in vitro [33]. Adipocytokine release (including tumor necrosis factor— α and PAI-1) was recently shown to be regulated in part by angiotensin II [34]. Thus, angiotensin II may trigger a cascade of cellular effects that results in amplified bFGF production and release from multiple storage sites.

The limitations of this study are that it is cross-sectional and the findings are applicable only to men. More study is needed in women with diabetes and in other racial and ethnic groups to determine whether differences in log plasma bFGF may mirror other population differences in PAI-1 concentration [20]. Still unexplained is whether unknown factors related to poor baseline glycemic control may have accounted for an unexpected inverse correlation between log bFGF and HbA_{1c} (Table 4), an association that could not be accounted for (in the multiple regression analysis) by race or waist-hip ratio.

In conclusion, plasma bFGF was unexpectedly elevated in a subgroup of non-Hispanic white patients with advanced type 2 diabetes mellitus. These findings suggest the possibility that markedly increased plasma bFGF may reflect not only impaired fibrinolysis in type 2 diabetes mellitus but also, consistent with its role as a broad-spectrum cellular mitogen, the promotion of atherosclerosis, and/or plaque neovascularization [35]. Whether log plasma bFGF or PAI-1 may be a better predictor of newly occurring atherothrombotic cardiovascular events in adult men with advanced type 2 diabetes mellitus can be tested in the ongoing VADT.

Acknowledgment

We thank Dr Carlos Abraira and Dr William Duckworth, Co-Chairmen of the VADT, for their encouragement and critical review of the manuscript.

Supported by a grant from the Veterans Biomedical Research Institute, East Orange, NJ (to MBZ), and by the Cooperative Studies Program of the Department of Veterans Affairs, Office of Research and Development, Washington, DC. The authors report no conflicts of interest that would affect the objectivity of the findings presented.

References

- Gospodarowicz D, Ferrara N, Schweigerer L, Neufeld G. Structural characterization and biological functions of fibroblast growth factor. Endocr Rev 1987;8:95-114.
- [2] Esch F, Baird A, Ling N, Ueno N, Hill F, Denoroy L, et al. Primary structure of bovine pituitary basic fibroblast growth factor (FGF) and comparison with the amino-terminal sequence of bovine brain acidic FGF. Proc Nat Acad Sci U S A 1985;19:6507-11.
- [3] Hasdai D, Barak V, Leibovitz E, Herz I, Sclarovsky S, Eldar M, et al. Serum basic fibroblast growth factor levels in patients with ischemic heart disease. Int J Cardiol 1997;59:133-8.
- [4] Katinioti AA, Tousoulis D, Economou E, Stefanadis C, Trikas A, Tentolouris C, et al. Basic fibroblast growth factor changes in response to coronary angioplasty in patients with stable angina. Int J Cardiol 2002;84:195-9.
- [5] Kraemer R, Pomerantz KB, Joseph-Silverstein J, Hajjar DP. Induction of basic fibroblast growth factor mRNA and protein synthesis in smooth muscle cells by cholesteryl ester enrichment and 25hydroxycholesterol. J Biol Chem 1993;268:8040-5.
- [6] Cucina A, Scavo MP, Muzzioli L, Coluccia P, Ceccarini S, Fuso A, et al. High density lipoproteins downregulate basic fibroblast growth factor production and release in minimally oxidated-LDL treated smooth muscle cells. Atherosclerosis 2006;189:303-9.
- [7] Abraira C, Duckworth W, McCarren M, Emanuele N, Arca D, Reda D, et al. Design of the cooperative study of glycemic control and complications in diabetes mellitus type 2. J Diabet Complications 2003;17:314-22.
- [8] Schweigerer L, Neufeld G, Friedman J, Abraham JA, Fiddes JC, Gospodarowicz D. Capillary endothelial cells express basic fibroblast growth factor, a mitogen that promotes their own growth. Nature 1987; 325:257-9.
- [9] Zimering MB, Eng J. Increased basic fibroblast growth factor—like substance in plasma from a subset of middle-aged or elderly male diabetic patients with microalbuminuria or proteinuria. J Clin Endocrinol Metab 1996;81:4446-52.
- [10] Zimering MB. Effect of intravenous bisphosphonates on release of basic fibroblast growth factor in serum of patients with cancerassociated hypercalcemia. Life Sci 2002;70:1-14.

- [11] Larsson A, Skoldenberg E, Ericson H. Serum and plasma levels of FGF-2 and VEGF in healthy blood donors. Angiogenesis 2002;5: 107-10.
- [12] Dirix LY, Vermeulen PB, Pawinski A, Prové A, Benoy I, De Pooter C, et al. Elevated levels of the angiogenic cytokines basic fibroblast growth factor and vascular endothelial growth factor in sera of cancer patients. Br J Cancer 1997;76:238-43.
- [13] Tabone MD, Landman-Parker J, Arcil B, Coudert MC, Gerota I, Benbunan M, et al. Are basic fibroblast growth factor and vascular endothelial growth factor prognostic indicators in pediatric patients with malignant solid tumors? Clin Cancer Res 2001;7:538-43.
- [14] Folkman J, Klagsbrun M. Angiogenic factors. Science 1987;235: 442-7
- [15] Kaneko T, Fujii S, Matsumoto A, Goto D, Ishimori N, Watano K, et al. Induction of plasminogen activator inhibitor—1 in endothelial cells by basic fibroblast growth factor and its modulation by fibric acid. Arterioscler Thromb Vasc Biol 2002;22:855-60.
- [16] Sahni A, Sahni SK, Simpson-Haidaris PJ, Francis CW. Fibrinogen binding potentiates FGF-2 but not VEGF induced expression of u-PA, u-PAR, and PAI-1 in endothelial cells. J Thromb Haemost 2004;2: 1629-36
- [17] Keber I, Keber D. Increased plasminogen activator inhibitor activity in survivors of myocardial infarction is associated with metabolic risk factors of atherosclerosis. Haemostasis 1992;22:187-94.
- [18] Thogersen AM, Jansson JH, Boman K, Nilsson TK, Weinehall L, Huhtasaari F, et al. High plasminogen activator inhibitor and tissue plasminogen activator levels in plasma precede a first acute myocardial infarction in both men and women: evidence for the fibrinolytic system as an independent primary risk factor. Circulation 1998;98:2241-7.
- [19] Wiman B, Andersson T, Hallqvist J, Reuterwall C, Ahlbom A, deFaire U. Plasma levels of tissue plasminogen activator/plasminogen activator inhibitor—1 complex and von Willebrand factor are significant risk markers for recurrent myocardial infarction in the Stockholm Heart Epidemiology Program (SHEEP) study. Arterioscler Thromb Vasc Biol 2000;20:2019-23.
- [20] Lutsey PL, Cushman M, Steffen LM, Green D, Barr RG, Herrington D, et al. Plasma hemostatic factors and endothelial markers in four racial/ ethnic groups: the MESA study. J Thromb Haemost 2006;4:2629-35.
- [21] Frist ST, Taylor Jr HA, Kirk KA, Grammer JR, Li XN, Grenett HE, et al. Expression of PAI-1, t-PA and u-PA in cultured human umbilical vein endothelial cells derived from racial groups. Thromb Res 1995; 77:279-90.
- [22] Alessi MC, Bastelica D, Morange P, Berthet B, Leduc I, Verdier M, et al. Plasminogen activator inhibitor 1, transforming growth factor–beta1, and BMI are closely associated in human adipose tissue during morbid obesity. Diabetes 2000;49:1374-80.
- [23] Flugelman MY, Virmani R, Correa R, Yu ZX, Farb A, Leon MB, et al. Smooth muscle cell abundance and fibroblast growth factors in coronary lesions of patients with nonfatal unstable angina. A clue to the mechanism of transformation from the stable to the unstable clinical state. Circulation 1993;88:2493-500.
- [24] Mydlo JH, Kral JG, Macchia RJ. Differences in prostate and adipose tissue basic fibroblast growth factor: analysis of preliminary results. Urology 1997;50:472-8.
- [25] Mydlo JH, Kral JG, Macchia RJ. Preliminary results comparing the recovery of basic fibroblast growth factor (FGF-2) in adipose tissue and benign and malignant renal tissue. J Urol 1998;159:2159-63.
- [26] Teichert-Kuliszewska K, Hamilton BS, Deitel M, Roncari DA. Augmented production of heparin-binding mitogenic proteins by preadipocytes from massively obese persons. J Clin Invest 1992;90: 1226-31.
- [27] Fain JN. Release of interleukins and other inflammatory cytokines by human adipose tissue is enhanced in obesity and primarily due to the nonfat cells. Vitam Horm 2006;74:443-77.
- [28] Liuzzo JP, Petanceska SS, Moscatelli D, Devi LA. Inflammatory mediators regulate cathepsin S in macrophages and microglia: a role in attenuating heparan sulfate interactions. Mol Med 1999;5:320-33.

- [29] Itoh H, Mukoyama M, Pratt R, Gibbons G, Dzau V. Multiple autocrine growth factors modulate vascular smooth muscle cell growth response to angiotensin II. J Clin Invest 1993;91:2268-74.
- [30] Alderman MH, Cohen HW, Sealey JE, Laragh JH. Plasma renin activity levels in hypertensive persons: their wide range and lack of suppression in diabetic and in most elderly patients. Am J Hypertens 2004;17:1-7.
- [31] Holland OB, von Kuhnert L, Campbell WB, Anderson RJ. Synergistic effect of captopril with hydrochlorothiazide for the treatment of lowrenin hypertensive black patients. Hypertension 1983;5:235-9.
- [32] Coppack SW. Pro-inflammatory cytokines and adipose tissue. Proc Nutr Soc 2001;60:349-56.
- [33] Samaniego F, Markham PD, Gendelman R, Gallo RC, Ensoli B. Inflammatory cytokines induce endothelial cells to produce and release basic fibroblast growth factor and to promote Kaposi's sarcoma-like lesions in nude mice. J Immunol 1997;158:1887-94.
- [34] Kurata A, Nishizawa H, Kihara S, Maeda N, Sonoda M, Okada T, et al. Blockade of angiotensin II type 1 receptor reduces oxidative stress in adipose tissue and ameliorates adipocytokine dysregulation. Kidney Int 2006;70:1717-24.
- [35] Hughes SE, Crossman D, Hall PA. Expression of basic and acidic fibroblast growth factors and their receptor in normal and atherosclerotic human arteries. Cardiovasc Res 1993;27:1214-9.